



## **PRENATAL EXPOSURE TO ENVIRONMENTAL CONTAMINANTS AND CHILD BEHAVIOUR IN THE HELIX EUROPEAN COHORTS**

**Paulina Jedynak**

Master's Degree in Bioinformatics and Biostatistics UOC-UB

Clinical Data Analysis

Professional supervisor:

**Claire Philippat**

Academic supervisor:

**Miguel-Ángel Mayer**

Submission date:

December 2018



This work is licensed under  
Attribution-NonCommercial-NoDerivs License  
[3.0 Spain Creative Commons](https://creativecommons.org/licenses/by-nc-nd/3.0/es/)

### THESIS WORK SHEET

<b>Title:</b>	<i>Prenatal exposure to environmental contaminants and child behaviour In the HELIX European cohorts</i>
<b>Author:</b>	<i>Paulina Jedynak</i>
<b>Professional supervisor:</b>	<i>Claire Philippat</i>
<b>Academic supervisor:</b>	<i>Miguel-Ángel Mayer</i>
<b>Submission date:</b>	<i>12/2018</i>
<b>Degree:</b>	<i>Master's Degree in Bioinformatics and Biostatistics UOC-UB</i>
<b>Area:</b>	<i>Clinical Data Analysis</i>
<b>Language:</b>	<i>English</i>
<b>Key words:</b>	<i>Prenatal exposure, Environmental contaminants, Child behaviour</i>

**Abstract:**

<p>=====</p> <p>Abstract was removed</p> <p>=====</p>
---

## TABLE OF CONTENTS

<i>THESIS WORK SHEET</i> .....	<i>i</i>
<i>LIST OF FIGURES</i> .....	<i>iii</i>
<i>LIST OF TABLES</i> .....	<i>iii</i>
<i>I. Introduction</i> .....	<i>1</i>
<i>1. Context and justification of the project</i> .....	<i>1</i>
<i>1.1. Child neuropsychological development</i> .....	<i>1</i>
<i>1.2. Risk factors for neurodevelopmental disorders</i> .....	<i>1</i>
<i>1.3. Environmental contaminants and neuropsychological deficits</i> .....	<i>2</i>
<i>1.3.1. Personal exposome concept</i> .....	<i>5</i>
<i>2. Objectives of the project</i> .....	<i>5</i>
<i>3. Materials &amp; Methods</i> .....	<i>5</i>
<i>3.1. Study population</i> .....	<i>5</i>
<i>3.2. Behavioural outcomes</i> .....	<i>8</i>
<i>3.2.1. "Externalising" and "Internalising" scores</i> .....	<i>8</i>
<i>3.3. Estimation of prenatal exposure to environmental chemicals</i> .....	<i>8</i>
<i>3.3.1. Dealing with missing exposure values</i> .....	<i>8</i>
<i>3.3.2. Dealing with undetected concentrations</i> .....	<i>9</i>
<i>3.3.3. Adjustment for lipids and creatinine</i> .....	<i>9</i>
<i>3.3.4. Transformations</i> .....	<i>9</i>
<i>3.4. Confounders</i> .....	<i>9</i>
<i>3.4.1. Coding of confounders</i> .....	<i>9</i>
<i>3.4.2. Dealing with missing confounder values</i> .....	<i>10</i>
<i>3.5. Statistical Methods</i> .....	<i>10</i>
<i>3.6. Correction for multiple comparisons</i> .....	<i>10</i>
<i>3.7. Sensitivity analysis</i> .....	<i>10</i>
<i>3.8. Statistical software</i> .....	<i>10</i>
<i>4. Project scheduling</i> .....	<i>10</i>
<i>5. Summary of the results</i> .....	<i>13</i>
<i>6. Summary of the chapters</i> .....	<i>13</i>
<i>V. Abbreviations and acronyms list</i> .....	<i>29</i>
<i>VI. Bibliography</i> .....	<i>30</i>

**LIST OF FIGURES**

*Figure 1:*----- 1  
*Figure 2:*----- 7  
*Figure 3:*----- 7

**LIST OF TABLES**

*Table 1:*----- 3  
*Table 2:*----- 6  
*Table 3:*----- 9

# I. Introduction

## 1. Context and justification of the project

### 1.1. Child neuropsychological development

Brain development begins in the womb, with most of the structural features of the brain formed by the eighth week of conception. These structures continue to grow and develop throughout pregnancy and after birth [1] and it is accepted that the developing brain (in foetal life and early childhood) is more vulnerable to injuries caused by environmental chemicals than the brain of an adult [2], [3]. Healthy neurodevelopment manifests with gradual achievement of psychomotor, cognitive and socio-emotional functions that enable a child to tackle different developmental challenges (see Figure 1). Deficits in any of these domains may cause clinical neurodevelopmental disorders disturbing personal, social, academic or occupational functioning later in life [4]. Disorders of neurobehavioral development affect 10–15% of all births [5] and the most prevalent conditions with onset in the developmental period are: communication impairments, specific learning and motor disorders, intellectual disability, attention-deficit hyperactivity disorder (ADHD) and autism spectrum disorder (ASD) [6], [7]. More recently described impairments, such as disruptive mood and conduct disorders, receive growing attention as potentially also rooting from neurodevelopmental deficiencies [8]. In the past years the prevalence of such disorders as ADHD and ASD has been increasing alarmingly [9] creating a need to pinpoint potential risk factors for such impairments to improve their treatment and prevention.

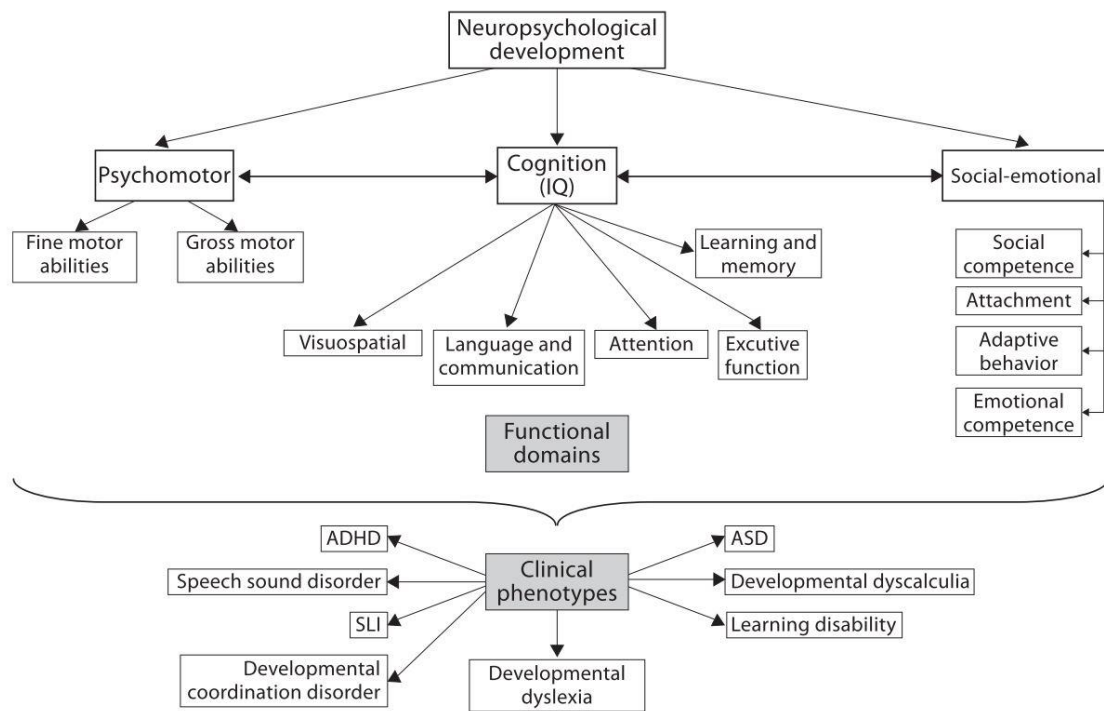


Figure 1: Conceptual framework of the neuropsychological development process. Abbreviations: SLI - specific language impairment; ADHD - attention deficit hyperactivity disorder; ASD - autistic spectrum disorder. Reprinted from Ref. [10]

### 1.2. Risk factors for neurodevelopmental disorders

Root causes of the most common early childhood neurodevelopmental disorders are multifactorial and only partly understood. Research on developmental biology and human and animal physiology showed that, apart from genetic factors, the environment of foetal and early childhood have strong effects on development, health maintenance, and

incidents of disease [11] which is the concept underlying the Developmental Origin of Health and Disease (DOHaD, [12]). Environmental factors that may play a significant role in vulnerability to neurodevelopmental disorders, particularly ASD and ADHD whose heritability was estimated to be around 50% [13]. The complementing half include maternal stress, some infections during pregnancy, malnutrition and maternal exposure to environmental toxicants [14], [15]. Approximately three percent of the developmental disabilities might be the direct result of pre- and postnatal exposure to environmental contaminants [16] that are ubiquitously present in our life, e.g. in food, drinking water or air.

### **1.3. Environmental contaminants and neuropsychological deficits**

Several groups of chemical compounds that are ubiquitously present in the environment were proved or suggested to have adverse neurodevelopmental effects on humans, manifested by behavioural, cognitive or motor deficits (Table 1). According to the most recent reviews of environmental agents affecting human neurodevelopment, a wide range of environmental chemicals, such as organochlorine compounds (OCs), polybrominated diphenyl ethers (PBDEs), perfluoroalkyl substances (PFASs), phthalates, bisphenol A, organophosphorus pesticides (OP Pesticides), metals as well as tobacco smoke are proved to disrupt endocrine function [17]–[20]. Endocrine disrupting chemicals (EDCs) are substances that may interfere with the body's endocrine system and thereby produce adverse health effects, including developmental, reproductive, neurological and immune effects in humans and animal models [21]. EDCs interfere, temporarily or permanently, with hormonal signalling pathways in the endocrine system. One of the EDCs' targets is thyroid signalling pathway [17], [22], [23], whose undisturbed functioning is crucial in normal brain development, including neural differentiation and migration and neural connectivity [24], [25]. Recent advances in exposure assessment and biostatistics allow a new approach in epidemiological studies in which effects of exposures to a broad spectrum of environmental contaminants from multiple sources could be evaluated in concert.

Table 1:  
Chemical compounds studied and examples of their environmental sources

Exposure family	Exposure	Acronym	Exemplary use/source	Adverse effect on neuropsychological development	
				Original study	Review
OCs	Dichlorodiphenyldichloroethylene	DDE	Food contaminated with pesticides	[26]	
	Dichlorodiphenyltrichloroethane	DDT		[27]	
	Hexachlorobenzene	HCB		[28]	
	Polychlorinated biphenyl-118	PCB-118	Transformers and capacitors, electrical equipment, industrial oils, cable and thermal insulation materials, adhesives and tapes, some plastics		
	Polychlorinated biphenyl-138	PCB-138			
	Polychlorinated biphenyl-153	PCB-153		[26], [29]	[30]
	Polychlorinated biphenyl-170	PCB-170			
Polychlorinated biphenyl-180	PCB-180				
PBDEs	Polybrominated diphenyl ether-47	PBDE-47	Flame retardants in textiles, plastics, wire insulation, automobiles	[31], [32]	[18]
	Polybrominated diphenyl ether-153	PBDE-153			
PFASs	Perfluorononanoate	PFNA	Food packages, stain- and water-repellent fabrics, non-stick products (e.g., Teflon), polishes, waxes, paints, cleaning products	Unequivocal adverse effects not proved: [33], [34]	[35]
	Perfluorohexane sulfonate	PFHXS			
	Perfluorooctanoate	PFOA			
	Perfluorooctane sulfonate	PFOS			
	Perfluoroundecanoic acid	PFUnDA			
Phthalates	Mono benzyl phthalate	MBzP	Plasticizers, solvents for other materials, food containers, vinyl flooring, adhesives, detergents, lubricating oils, automotive plastics, plastic clothes (raincoats), and personal-care products (soaps, shampoos, hair sprays, and nail polishes)	[36]–[43]	[44]
	Mono-2-ethyl-5-carboxypentyl phthalate	MECPP			
	Mono-2-ethyl-5-hydroxyhexyl phthalate	MEHHP			
	Mono-2-ethylhexyl phthalate	MEHP			
	Mono-2-ethyl-5-oxohexyl phthalate	MEOHP			
	Monomethyl phthalate	MEP			
	Mono-iso-butyl phthalate	MiBP			
	Mono-n-butyl phthalate	MnBP			
	Mono-4-methyl-7-hydroxyoctyl phthalate	ohMiNP			
Mono-oxo-isononyl phthalate	oxo-MiNP		Effect not studied		



	Bisphenol A	BPA	Polycarbonate containers and coatings (cans, cups), dental sealants, cash register receipts	[45]–[51]	
Phenols	Parabens: N-Butyl paraben, Ethyl paraben, Methyl paraben, Propyl paraben	BUPA, ETPA, MEPA, PRPA	Preservatives in cosmetics like makeup, moisturizers, hair care products, shaving products and food products to prevent spoilage	Adverse effect not proved: [43], [53]	[52]
	Oxybenzone	OXBE	Sunscreens		
	Triclosan	TRCS	Microbicide in home cleaning and personal care products, anti-bacterial soaps and toothpaste	[43] Adverse effect not proved: [54]	
OP Pesticides	Diethyl phosphate	DEP			
	Diethyl thiophosphate	DETP			
	Dimethyl dithiophosphate	DMDTP	Pesticides including chloropyrifos, parathion, malathion and diazinon	[55]–[57]	[58]
	Dimethyl phosphate	DMP			
	Dimethyl thiophosphate	DMTP			
Metals	Arsenic, Cadmium, Cobalt, Caesium, Copper, Mercury, Potassium, Magnesium, Manganese, Molybdenum, Sodium, Lead, Selenium, Zinc	As, Cd, Co, Cs, Cu, Hg, K, Mg, Mn, Mo, Na, Pb, Se, Zn	Industrial contamination, electronic devices, pigments, batteries, plastics, tobacco smoke, preservatives, combustion fumes, fertilizers, and many more	[59]–[64]; Unequivocal adverse effects not proved: As, Hg, Co, Mg, Tl. Effect not studied: Cu, Cs, K, Na	[22], [65]
	Cotinine		Tobacco smoke	[66]–[68]	[69]

Abbreviations: OCS - organochlorine compounds; PBDEs - polybrominated diphenyl ethers; PFASs - perfluoroalkyl substances; OP Pesticides - organophosphorus pesticides

### **1.3.1. Personal exposome concept**

Pregnant women from the general population are exposed to a broad range of environmental chemicals some of which can cross the placental barrier and affect the foetus [70], [71]. Until now, epidemiological studies on the effects of environmental contaminants on child neurodevelopment usually consider each exposure (or family of exposures) separately, and therefore provide only a fragmented view of environment-health associations. New studies that would simultaneously take into account multiple exposures were needed to improve the understanding of the environmental risk factors for neurodevelopmental disorders. The personal exposome concept, defined as the totality of non-genetic exposures from conception throughout the life course [72], comprises a wide range of environmental exposures from the prenatal period onwards, providing a holistic consideration of many exposures simultaneously which in turn helps to understand the complex environmental component of disease aetiology. One of the first epidemiological study implementing the personal exposome concept is the European human early life exposome (HELIX) project [73]. The aim of the HELIX project was to measure and describe exposure to a multitude of environmental contaminants during early life (pregnancy and childhood) in 6 prospective European cohorts and associate these exposures with child health outcomes [71], [74]. Such approach derives from the exposome concept and aims at estimating effect of combined exposures (e.g., based on biological pathways) on health and are likely to be more realistic than the classical approach focused on each chemical separately.

## **2. Objectives of the project**

The focus of this study was to assess the associations between prenatal exposure to a wide range of environmental chemicals, including some that have been identified as neurodevelopmental disruptors, and child externalising and internalising behaviour between three and seven years of age in the European HELIX cohorts.

## **3. Materials & Methods**

### **3.1. Study population**

The study population consisted of 1321 mother-child pairs selected from the Human Early Life Exposome (HELIX) study [73]. The HELIX cohorts consist of 31 472 pregnant women recruited during pregnancy between 2003 and 2010, in the six European countries: EDEN (France), RHEA (Greece), KANC (Lithuania), MoBa (Norway), INMA (Spain) and BiB (the UK). The characteristics of the sub-cohorts contributing to the HELIX cohort are presented in the Table 2 [75].

Table 2:

The characteristics of the sub-cohorts contributing to the HELIX project. Reprinted from Ref. [75]

Cohort	Recruitment in original cohort	Exclusions made during recruitment	Years of birth	Region covered by HELIX	No. of births in HELIX entire cohort
BiB, UK <sup>5</sup>	All pregnant women who attended the oral glucose tolerance test clinic at Bradford Royal Infirmary in weeks 26–28 of pregnancy.	Women who planned to move away from Bradford before birth were excluded.	2007–2010	Bradford	10 849
EDEN, France <sup>6</sup>	Pregnant women who attended prenatal care at the University hospitals of Nancy and Poitiers recruited before 24 weeks of amenorrhoea.	Twin pregnancies, women with known diabetes before pregnancy, insufficient French language skills and intention to move away from the recruitment area were excluded.	2003–2006	Nancy and Poitiers, urban areas	1900
INMA, Spain <sup>7</sup>	Pregnant women who attended a prenatal care centre in the study region during weeks 6–10 of pregnancy.	Women who resided or intended to deliver outside the study area, who were aged under 16 years, who had twin or multiple pregnancies, who had assisted reproduction or who had communication problems were excluded.	2003–2008	Gipuzkoa Sabadell Valencia	2063
KANC, Lithuania <sup>8</sup>	Pregnant women who attended one of four prenatal care clinics affiliated to the hospitals of the Kaunas University of Medicine during first trimester of pregnancy.	Women who lived outside Kaunas municipality, had medical records of pregnancy induced hypertension and/or diabetes were excluded.	2007–2008	Kaunas	4107
MoBa, Norway <sup>9</sup>	Recruitment at the first ultrasound (US) scan, ie, during the 17–18 weeks of gestation. All women who gave singleton births in the participating maternity units.	None	1999–2008	Oslo	11 095
RHEA, Greece <sup>10</sup>	Pregnant women who attended US examination before 15 week of pregnancy with residence in and near Heraklion at Crete.	Women who were aged under 16 years or who had communication problems were excluded.	2007–2008	Heraklion	1458
Total					31 472

BiB, Born in Bradford; EDEN, Étude des Déterminants pré et postnatals du développement et de la santé de l'Enfant; INMA, Infancia y Medio Ambiente; KANC, Kaunas cohort; MoBa, Norwegian Mother and Child Cohort Study.

The study design is depicted in Figure 2. For a sub-cohort of 1321 mother-child pairs, biomarkers of exposure to several contaminants were measured in pregnant women's blood and urine. Among them, 708 had behavioural outcomes of their children assessed between three to seven years of age [74] using the Strengths and Difficulties Questionnaire (SDQ) [76]. Inclusion criteria are illustrated in Figure 3.

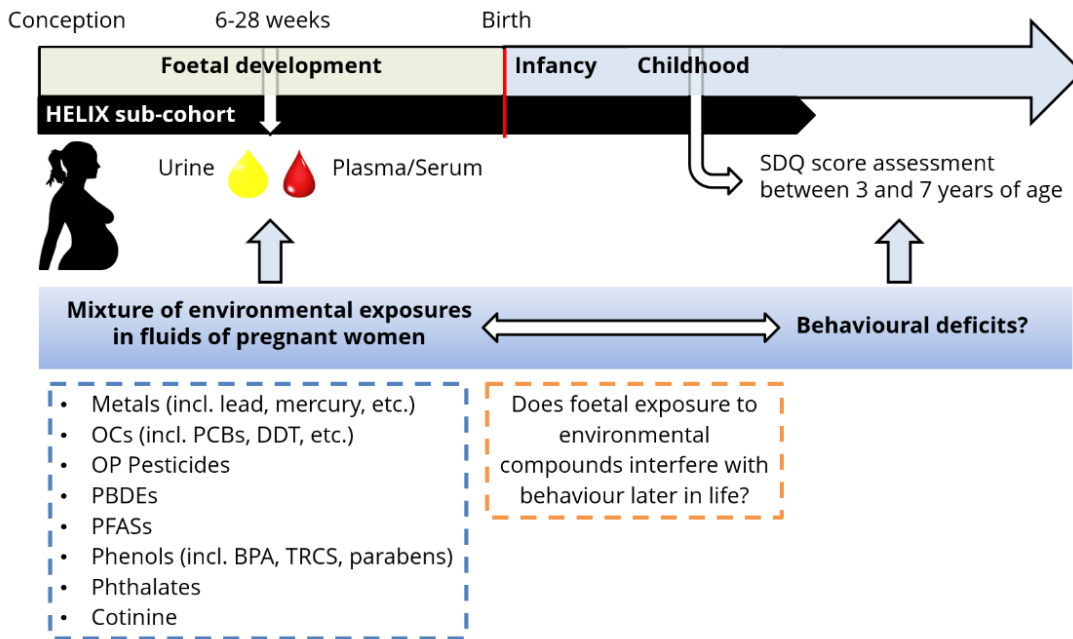


Figure 2:  
Study design for the HELIX cohorts

**HELIX original study population (N = 1321)**

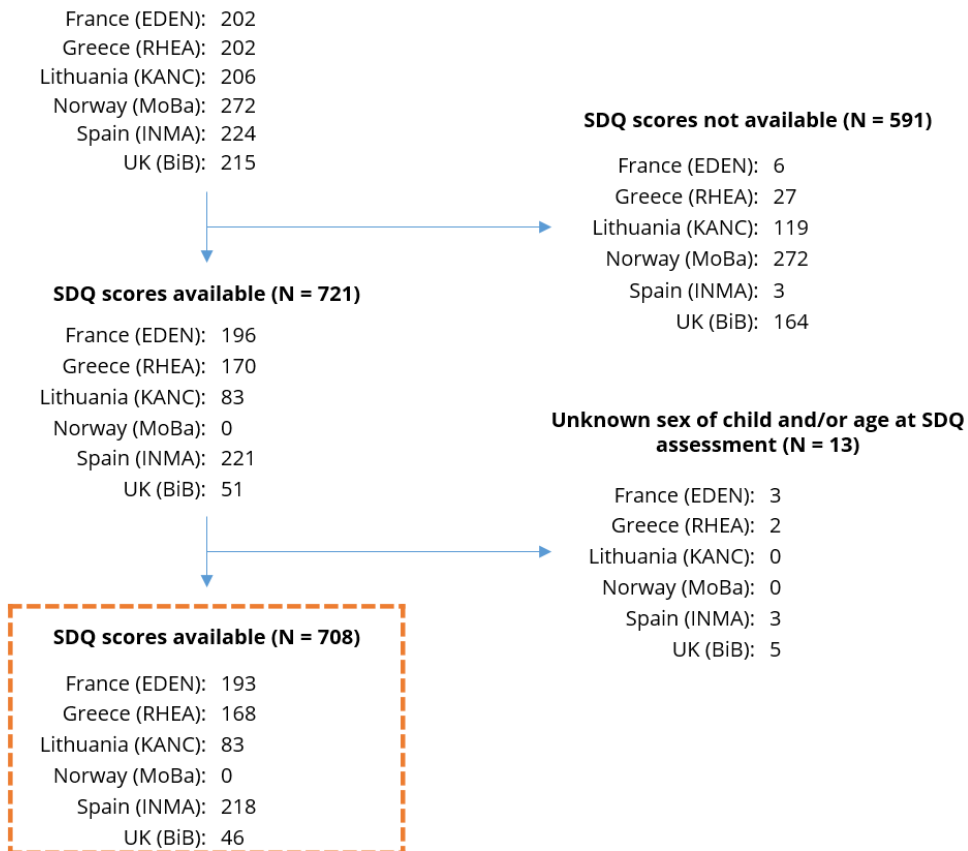


Figure 3:  
Flowchart illustrating selection of the study population from the HELIX cohorts

### 3.2. Behavioural outcomes

Behaviour was assessed using the Strengths and Difficulties Questionnaire (SDQ) [76] between three and seven years of age. The SDQ consists of the 25 items that were combined into four difficulties sub-scores: conduct problems, hyperactivity–inattention, peer relationship problems and emotional symptoms sub-scores and one strength sub-score: prosocial behaviour. Each item can be evaluated as “Not True”, “Somewhat True” and “Certainly True”. “Somewhat True” is always scored as 1, but the scoring of “Not True” and ‘Certainly True’ varies with the item. For each of the 5 scales the score can range from 0 to 10 if all items were completed. Higher scores on the behavioural scales indicate increased behaviour problems.

#### 3.2.1. “Externalising” and “Internalising” scores

The externalising score is the sum of the conduct and hyperactivity–inattention sub-scores and ranges from 0 to 20 and. The internalising score also ranges from 0 to 20 and is the sum of the emotional and peer problems sub-scores. Using these two amalgamated scales may be preferable to using the four separate scales in community samples [77] and therefore were used in the present study.

### 3.3. Estimation of prenatal exposure to environmental chemicals

We assessed 52 environmental contaminants that are ubiquitously present in our lives. We added one grouping variable consisting of the molar sum of 4 di(2-ethylhexyl) phthalate (DEHP) metabolites ( $\Sigma$ DEHP). The studied compounds came from the following groups: organochlorine compounds (OCs), polybrominated diphenyl ethers (PBDEs), perfluoroalkyl substances (PFASs), phthalates, phenols, organophosphorous pesticides (OP Pesticides), metals, and cotinine - a marker of smoking. OCs, PFASs PBDEs and metals are long half-life chemicals and were measured in maternal blood while concentrations of compounds with short half-life (OP Pesticides, phenols and phthalates) were assessed in maternal urine. The maternal samples from the various cohorts were collected at different time points [71]; INMA, KANC, RHEA recruited during the first trimester, EDEN recruited during the first and second trimesters and BIB recruited during the second and third trimesters. The mean ( $\pm$  SD) gestational ages (weeks) at blood sample collection were 26.6 ( $\pm$  1.4), 26.1 ( $\pm$  1.2), 13.7 ( $\pm$  2.0), 39.4 ( $\pm$  1.3) and 14.1 ( $\pm$  3.7) for BIB, EDEN, INMA, KANC and RHEA, respectively. Urine sample collection times were the same as for blood for BIB, EDEN and RHEA cohorts. In INMA cohort urine was collected at mean ( $\pm$  SD) gestational age (weeks) of 34.2 ( $\pm$  1.3) while for KANC cohort urine samples were not collected at all. For more details of samples processing please refer to the HELIX project please refer to Huag *et al.* In Table 1 (see “Introduction, 1.3 Environmental contaminants and neuropsychological deficits”) the studied exposure biomarkers are listed together with examples of their environmental sources.

#### 3.3.1. Dealing with missing exposure values

For some sub-cohorts and/or participants within a sub-cohort, blood or urine samples were missing preventing us from measuring some biomarkers of exposure. As it has been shown that multiple imputation is superior to excluding a variable or a stratum [78], [79], exposures for which less than 70% of observations were missing due to not being measured were imputed using multiple imputation (20) relying on the method of chained equations algorithm (*mice* R package) [80], [81]. The algorithms used for the imputation were the following: logistic regression for binomial variables; predictive mean matching for continuous variables; polytomous logistic regression for multinomial variables. The imputation predictors list was limited to only those variables that were not highly correlated with other variables to prevent imputation errors due to multicollinearity. The condition for variable removal from the predictors list was based on the number of variables it was highly correlated with (Spearman corr. coefficient  $\rho \geq 0.6$ ). If there was more than 1 variable with the same number of correlated partners, variable with higher number of complete cases number was favoured. Highly correlated variables were removed from prediction of all other imputed variables and not only from the imputation of their correlated counterparts (as such approach produced better convergence plots). An additional restriction of the predictor list was applied based on the influx/outflux plot [82]. Variables with

low predictive ability (outflux < 0.5) were removed. Passive imputation was applied to impute transformed (e.g. log2) or combined (e.g. sum DEHP) versions of the data [83]. Diagnostics was performed at the end of the imputation process in order to check for imputation convergence and plausibility of the newly generated values (see **Error! Reference source not found.** and **Error! Reference source not found.**).

### 3.3.2. Dealing with undetected concentrations

Exposures for which more than 80% of observations were below the limit of detection (LOD) were dichotomized into detected/undetected, as advised by the HELIX consortium. For the other variables, values below the LOD were singly imputed using a quantile regression approach for the imputation of left-censored missing data.

### 3.3.3. Adjustment for lipids and creatinine

To limit the impact of between-subject variations in urine and blood dilution, the biomarker concentrations were adjusted for creatinine concentration (urine) or total fat percentage (blood) [84], [85]. This adjustment was done by dividing the urine-based biomarker concentrations by the creatinine concentrations and serum-based biomarker concentrations by total fat percentage. Chemicals from the PFASs family and metals, although collected from blood, were not adjusted for lipids as they are not lipophilic.

### 3.3.4. Transformations

The continuous biomarker concentrations were log-transformed (base 2) prior to analysis to limit the effect of outliers.

## 3.4. Confounders

A confounder is defined as variable that is associated with both the exposure and the outcome but that is not a consequence of either [86]. Information on potential confounders such as maternal behaviours and socio-economic status was collected by interview during study visits and self-report questionnaires completed by the parents. Candidate confounders were selected based on literature review.

### 3.4.1. Coding of confounders

To establish the best coding for continuous confounders, all continuous adjustment factors were fitted in step functions to examine the relationship between the covariates and the outcomes (see **Error! Reference source not found.** for example of different types of coding of maternal Body Mass Index - BMI). The best coding was chosen for the confounder basing on the lower Akaike information criterion (AIC), graphical relationship between covariate and outcome and the degrees of freedom lost. Using this approach, we selected an appropriate coding for the confounders included in our multiple models as summarised in Table 3.

*Table 3:  
Confounders list*

Confounder group	Confounder	Variable type
Maternal behaviour	Smoking during pregnancy	Categorical (yes/ no)
	Breastfeeding	Categorical (yes/ no)
Socio-economic status	Maternal level of education	Categorical (primary school/ secondary school/ university degree or higher)
	Maternal work status	Categorical (yes/ no)
	Persons in household	Numerical
Other	Cohort origin	Categorical (France/ Greece/ Lithuania/ Spain/ UK)

Parity	Categorical (nulliparous / 1 child/ $\geq 2$ children)
Maternal age	Numerical
Maternal Body Mass Index	Categorical (underweight/ normal weight/ overweight/ obesity)
Child's sex	Categorical (male/ female)
Child's age at SDQ assessment	Numerical

### 3.4.2. Dealing with missing confounder values

As for the missing exposure concentrations, the missing values for confounder variables were imputed using multiple imputation based on the chained equations algorithm (*mice* R package), as described previously.

### 3.5. Statistical Methods

The outcome variables studied in the present project are behavioural SDQ scores that are non-negative, integer counts. Such data can possibly follow Poisson or negative binomial distribution. The negative binomial is a generalization of Poisson regression which loosens the restrictive assumption that the variance is equal to the mean made by the Poisson model [87]. In our case, the assumption of equality of the variance and the mean is not met and therefore we used the negative binomial regression model. In our study we performed separate adjusted negative binomial regressions to study the associations between each urinary or blood exposure concentration and the behavioural outcomes (SDQ scores). For SDQ scores, a positive effect estimate indicated an increased risk for behavioural problems while a negative effect estimate indicated a protective association. The effects were expressed as incidence rate ratios (IRR) for a doubling in biomarkers concentrations.

### 3.6. Correction for multiple comparisons

Since the simple regression was run for over 50 exposures a correction for multiple comparisons was applied to decrease the false discovery rate (FDR). As several of the explanatory variables were correlated we applied a modified Bonferroni p value adjustment according to an effective number of independent tests instead of the total tests number [88].

### 3.7. Sensitivity analysis

Good modelling practice requires providing an evaluation of the confidence in the proposed model [89]. In our sensitivity analysis we confronted the observed associations between exposures and SDQ scores in the imputed dataset with results obtained in complete case analysis to exclude the bias caused by the imputation process. Moreover, we run additional models with an interaction term between sex and exposure to investigate sex specific associations. Exposures whose interaction coefficients were significantly different from 1 (uncorrected p value  $< 0.2$ ) were analysed in separate simple regression analyses stratified by sex. Finally, to make sure that our results were not driven by the effect observed for one cohort, we run models based on the leave-one-cohort-out method and compared these results to our main analysis' outcomes. Exposures whose coefficients were significantly different from 1 in the main analysis (uncorrected p value  $< 0.05$ ) were re-analysed for subsets of data with one cohort removed at a time.

### 3.8. Statistical software

Statistical analyses were carried out using an open source software R version 3.5.1 and R-Studio version 1.1.456.

## 4. Project scheduling

The total time reserved for the experimental part of the project was approximately 9 weeks. Below an approximate timing for each part of the study is presented.

- 1) Described the levels of exposure to environmental contaminants among mother and children of the HELIX mother-child cohort (2 weeks)
  - a. Provided a descriptive summary of each environmental exposure (1 week)
    - Described the exposure variables – identified samples missing and below level of detection, described the distributions of the exposure concentrations, flagged potential outliers, measured correlations between exposures
  - b. Prepared the exposures data for statistical analyses (1 week)
    - Dichotomised exposures with too high number of samples whose concentration is below the level of detection
    - Transformed continuous exposures to normalize distributions
- 2) Identified covariates that could potentially influence exposure to environmental contaminants (2.5 weeks)
  - c. Selected candidate confounders (1 week)
    - Searched the literature in order to identify candidate confounders
    - Tested each potential confounder identified in the previous step in a simple regression against the behavioural outcome in order to establish the associations between the factors influencing exposure to environmental contaminants and behavioural score
  - d. Chose the confounders set and their optimal coding (1.5 weeks)
    - Tried different coding for the selected confounders (continuous, step function, etc.) and used AIC in order to establish the coding best reflecting the nature of the variable structure
    - Run multiple regression of different sets of confounders against the dependent variable in order to establish the configuration of confounders that describes best the variability of the behavioural outcome
- 3) Determined associations between the personal exposome and behavioural outcomes (4.5 weeks)
  - e. Pre-processed the final set of exposures, covariates and outcome variables (1 week)
    - Adjusted the exposure concentrations for urine and blood dilution using creatinine and total fat percentage, respectively
    - Imputed the missing values and/ or values below the limit of detection; Performed diagnostics of the imputation process and outcomes
  - f. Described associations between each exposure and the behavioural outcome (1.5 weeks)
    - Run exposome-wide association study (ExWAS) on outcome variable vs. each exposure adjusted for the confounders in order to establish the potential associations
  - g. Provided evaluation of the proposed model (1 week)
    - Run sensitivity analyses including factors that could possibly confound the observed associations between exposome and behavioural outcome

Below a graphical representation of the project scheduling is presented:



	2018								
	October			November				December	
	Week 1	Week 2	Week 3	Week 4	Week 5	Week 6	Week 7	Week 8	Week 9
	15-22/10	22-29/10	29/10-05/11	05-12/11	12-19/11	19-26/11	26/11-03/12	03-10/12	10-17/12
<b>1) Describe the levels of exposure to environmental contaminants among mother and children of the HELIX mother-child cohort</b> a. Provide a descriptive summary of each environmental exposure <ul style="list-style-type: none"> <li>Describe the exposure variables – identify samples missing and below level of detection, describe the distributions of the exposure concentrations, flag potential outliers, measure correlations between exposures</li> </ul>									
b. Prepare the exposures data for further analyses <ul style="list-style-type: none"> <li>If necessary, dichotomise exposures with too high number of samples whose concentration is below the level of detection</li> <li>If necessary, transform continuous exposures to normalize distributions and eliminate outliers</li> </ul>									
<b>2) Identify covariates that could potentially influence exposure to environmental contaminants</b> c. Select candidate confounders <ul style="list-style-type: none"> <li>Search the literature in order to identify candidate confounders</li> <li>Test each potential confounder identified in the previous step in a simple regression against the behavioural outcome in order to establish the associations between the factors influencing exposure to environmental contaminants and behavioural score</li> </ul>									
d. Choose the confounders set and their optimal coding <ul style="list-style-type: none"> <li>Try different coding for the selected confounders (continuous, step function, etc.) and use AIC in order to establish the coding best reflecting the nature of the variable structure</li> <li>Run multiple regression of different sets of confounders against the dependent variable in order to establish the configuration of confounders that describes best the variability of the behavioural outcome</li> </ul>									
<b>3) Determine associations between the personal exposome and behavioural outcomes</b> e. Pre-process the final set of exposures, covariates and outcome variables <ul style="list-style-type: none"> <li>Adjust the exposure concentrations for urine and blood dilution using creatinine and total, respectively</li> <li>Impute the missing values and/ or values below the limit of detection; Perform diagnostics of the imputation process and outcomes</li> </ul>									
f. Describe associations between each exposure and the behavioural outcome <ul style="list-style-type: none"> <li>Run exposome-wide association study (ExWAS) on outcome variable vs. each exposure adjusted for the confounders in order to establish the potential associations</li> </ul>									
g. Provide evaluation of the proposed model <ul style="list-style-type: none"> <li>Run sensitivity analysis including factors that could possibly confound the observed associations between exposome and behavioural outcome</li> </ul>									
h. <i>OPTIONALLY: Describe associations between a set of multiple exposures and the behavioural outcome</i> <ul style="list-style-type: none"> <li><i>Run elastic net regularized regression model (ENET) on a set of exposures against the dependent variable adjusted for the confounders in order to establish the associations between personal exposome and the behavioural outcome (with regard to false discovery rate correction)</i></li> </ul>									

## **5. Summary of the results**

We provided a detailed description of the studied population including maternal exposures and child behavioural outcomes; We identified confounders that could potentially influence exposure to environmental contaminants and behavioural outcomes; We described associations between prenatal exposures and postnatal behavioural outcomes; We validated the observed results by running several sensitivity analyses.

## **6. Summary of the chapters**

Chapter II – Results including the description of the studied population (part I) and of the associations between prenatal exposure to environmental contaminants and child behaviour (part II).

Chapter IV – Conclusions including the highlights of the present project and proposed approaches for future work.

Chapter V – Abbreviations and acronyms list including the terms used in the present work, in alphabetical order.

Chapter VI – Bibliography including the full list of works and websites consulted for this work.

Chapter VII – Annexes including results for the imputation diagnostics, example of comparison of different coding of a covariate, distribution of behavioural outcomes in the studied population, distribution of adjusted and transformed exposure concentrations, numerical summary of all simple regressions run on each exposure against behavioural outcome, and results of the sensitivity analyses.

=====

Pages 14 – 28 were removed

=====

## V. Abbreviations and acronyms list

<b>ADHD</b>	Attention-deficit hyperactivity disorder	<b>MEOHP</b>	Mono-2-ethyl-5-oxohexyl phthalate
<b>AIC</b>	Akaike information criterion	<b>MEP</b>	Monoethyl phthalate
<b>As</b>	Arsenic	<b>MEPA</b>	Methyl paraben
<b>ASD</b>	Autism spectrum disorder	<b>Mg</b>	Magnesium
<b>BMI</b>	Body mass index	<b>MiBP</b>	Mono-iso-butyl phthalate
<b>BPA</b>	Bisphenol A	<b>Mn</b>	Manganese
<b>BUPA</b>	N-Butyl paraben	<b>MnBP</b>	Mono-n-butyl phthalate
<b>CBCL</b>	Children behaviour checklist	<b>Mo</b>	Molybdenum
<b>Cd</b>	Cadmium	<b>Na</b>	Sodium
<b>CI</b>	Confidence interval	<b>OCs</b>	Organochlorine compounds
<b>Co</b>	Cobalt	<b>ohMINP</b>	Mono-4-methyl-7-hydroxyoctyl phthalate
<b>Cs</b>	Caesium	<b>OP Pesticides</b>	Organophosphorus pesticides
<b>Cu</b>	Copper	<b>OXBE</b>	Oxybenzone
<b>DDE</b>	Dichlorodiphenyldichloroethylene	<b>oxo-MINP</b>	Mono-4-methyl-7-oxooctyl phthalate
<b>DDT</b>	Dichlorodiphenyltrichloroethane	<b>Pb</b>	Lead
<b>DEHP</b>	Di-2-ethylhexyl-phthalate	<b>PBDE-153</b>	Polybrominated diphenyl ether-153
<b>DEP</b>	Diethyl phosphate	<b>PBDE-47</b>	Polybrominated diphenyl ether-47
<b>DETP</b>	Diethyl thiophosphate	<b>PBDEs</b>	polybrominated diphenyl ethers
<b>DMDTP</b>	Dimethyl dithiophosphate	<b>PCB-118</b>	Polychlorinated biphenyl-118
<b>DMP</b>	Dimethyl phosphate	<b>PCB-138</b>	Polychlorinated biphenyl-138
<b>DMTP</b>	Dimethyl thiophosphate	<b>PCB-153</b>	Polychlorinated biphenyl-153
<b>DOHaD</b>	Developmental origin of health and disease	<b>PCB-170</b>	Polychlorinated biphenyl-170
<b>EDCs</b>	Endocrine disrupting chemicals	<b>PCB-180</b>	Polychlorinated biphenyl-180
<b>ETPA</b>	Ethyl paraben	<b>PFASs</b>	Perfluoroalkyl substances
<b>FDR</b>	False discovery rate	<b>PFHXS</b>	Perfluorohexane sulfonate
<b>HCB</b>	Hexachlorobenzene	<b>PFNA</b>	Perfluorononanoate
<b>HELIX</b>	European human early life exposome	<b>PFOA</b>	Perfluorooctanoate
<b>Hg</b>	Mercury	<b>PFOS</b>	Perfluorooctane sulfonate
<b>IRR</b>	Incidence rate ratio	<b>PFUnDA</b>	Perfluoroundecanoate
<b>K</b>	Potassium	<b>PRPA</b>	Propyl paraben
<b>LOD</b>	Limit of detection	<b>SDQ</b>	Strengths and Difficulties Questionnaire
<b>MBzP</b>	Mono benzyl phthalate	<b>Se</b>	Selenium
<b>MECPP</b>	Mono-2-ethyl 5-carboxypentyl phthalate	<b>SLI</b>	Specific language impairment
<b>MEHHP</b>	Mono-2-ethyl-5-hydroxyhexyl phthalate	<b>TRCS</b>	Triclosan
<b>MEHP</b>	Mono-2-ethylhexyl phthalate	<b>Zn</b>	Zinc

## VI. Bibliography

- [1] A. S. Davis, *The handbook of pediatric neuropsychology*. Springer, 2011.
- [2] D. C. Bellinger, J. A. Matthews-Bellinger, and K. Kordas, "A developmental perspective on early-life exposure to neurotoxicants," *Environ. Int.*, vol. 94, pp. 103–112, Sep. 2016.
- [3] A. De Felice, L. Ricceri, A. Venerosi, F. Chiarotti, and G. Calamandrei, "Multifactorial Origin of Neurodevelopmental Disorders: Approaches to Understanding Complex Etiologies," *Toxics*, vol. 3, no. 4, pp. 89–129, 2015.
- [4] D. C. Bellinger, "Interpreting epidemiologic studies of developmental neurotoxicity: Conceptual and analytic issues," *Neurotoxicol. Teratol.*, vol. 31, no. 5, pp. 267–274, Sep. 2009.
- [5] B. Bloom, R. A. Cohen, and G. Freeman, "Summary health statistics for U.S. children: National Health Interview Survey, 2010," *Vital Health Stat. 10.*, no. 250, pp. 1–80, Dec. 2011.
- [6] American Psychiatric Association, *Diagnostic and Statistical Manual of Mental Disorders Fifth Edition (DSM-5)*. 2013.
- [7] H. D'Souza and A. Karmiloff-Smith, "Neurodevelopmental disorders," *Wiley Interdiscip. Rev. Cogn. Sci.*, vol. 8, no. 1–2, p. e1398, Jan. 2017.
- [8] C. D. Wills, "DSM-5 and neurodevelopmental and other disorders of childhood and adolescence.," *J. Am. Acad. Psychiatry Law*, vol. 42, no. 2, pp. 165–72, 2014.
- [9] P. J. Landrigan, L. Lambertini, and L. S. Birnbaum, "A Research Strategy to Discover the Environmental Causes of Autism and Neurodevelopmental Disabilities," *Environ. Health Perspect.*, vol. 120, no. 7, pp. a258–a260, Apr. 2012.
- [10] J. Fornis, A. Aranbarri, J. Grellier, J. Julvez, M. Vrijheid, and J. Sunyer, "A Conceptual Framework in the Study of Neuropsychological Development in Epidemiological Studies," *Neuroepidemiology*, vol. 38, no. 4, pp. 203–208, 2012.
- [11] P. D. Gluckman, "Living with the Past: Evolution, Development, and Patterns of Disease," *Science (80-. )*, vol. 305, no. 5691, pp. 1733–1736, Sep. 2004.
- [12] D. J. P. Barker, "The origins of the developmental origins theory," *J. Intern. Med.*, vol. 261, no. 5, pp. 412–417, May 2007.
- [13] S. Sandin, P. Lichtenstein, R. Kuja-Halkola, H. Larsson, C. M. Hultman, and A. Reichenberg, "The Familial Risk of Autism," *JAMA*, vol. 311, no. 17, p. 1770, May 2014.
- [14] E. Sciberras, M. Mulraney, D. Silva, and D. Coghill, "Prenatal Risk Factors and the Etiology of ADHD—Review of Existing Evidence," *Curr. Psychiatry Rep.*, vol. 19, no. 1, p. 1, Jan. 2017.
- [15] K. Lyall, R. J. Schmidt, and I. Hertz-Picciotto, "Maternal lifestyle and environmental risk factors for autism spectrum disorders," *Int. J. Epidemiol.*, vol. 43, no. 2, pp. 443–464, Apr. 2014.
- [16] P. Grandjean and P. J. Landrigan, "Neurobehavioural effects of developmental toxicity," *Lancet Neurol.*, vol. 13, no. 3, pp. 330–338, Mar. 2014.
- [17] A. Ghassabian and L. Trasande, "Disruption in Thyroid Signaling Pathway: A Mechanism for the Effect of Endocrine-Disrupting Chemicals on Child Neurodevelopment," *Front. Endocrinol. (Lausanne)*, vol. 9, no. APR, Apr. 2018.
- [18] N. Q. V. Tran and K. Miyake, "Neurodevelopmental Disorders and Environmental Toxicants: Epigenetics as an Underlying Mechanism," *Int. J. Genomics*, vol. 2017, pp. 1–23, 2017.
- [19] V. A. Rauh and A. E. Margolis, "Research Review: Environmental exposures, neurodevelopment, and child mental health - new paradigms for the study of brain and behavioral effects," *J. Child Psychol. Psychiatry*, vol. 57, no. 7, pp. 775–793, Jul. 2016.
- [20] M. Aschner and L. G. Costa, Eds., *Advances in neurotoxicology. Linking Environmental Exposure to Neurodevelopmental Disorders*. London: Academic Press, 2018.
- [21] D.-H. Lee, "Evidence of the Possible Harm of Endocrine-Disrupting Chemicals in Humans: Ongoing Debates and Key Issues," *Endocrinol. Metab.*, vol. 33, no. 1, p. 44, 2018.
- [22] K. von Stackelberg, E. Guzy, T. Chu, and B. C. Henn, "Exposure to Mixtures of Metals and Neurodevelopmental Outcomes: A Multidisciplinary Review Using an Adverse Outcome Pathway Framework," *Risk Anal.*, vol. 35, no. 6, pp. 971–1016, Jun. 2015.
- [23] T. T. Schug *et al.*, "Minireview: Endocrine Disruptors: Past Lessons and Future Directions," *Mol. Endocrinol.*, vol. 30, no. 8, pp. 833–847, Aug. 2016.
- [24] V. Somogyi *et al.*, "Bisphenol A influences oestrogen- and thyroid hormone-regulated thyroid hormone receptor

expression in rat cerebellar cell culture," *Acta Vet. Hung.*, vol. 64, no. 4, pp. 497–513, Dec. 2016.

- [25] World Health Organization, *State of the Science of Endocrine Disrupting Chemicals - 2012*. Geneva, 2013.
- [26] A. H. Rosenquist *et al.*, "Prenatal and Postnatal PCB-153 and p,p'-DDE Exposures and Behavior Scores at 5–9 Years of Age among Children in Greenland and Ukraine," *Environ. Health Perspect.*, vol. 125, no. 10, p. 107002, Oct. 2017.
- [27] F. W. Gaspar *et al.*, "Prenatal DDT and DDE exposure and child IQ in the CHAMACOS cohort," *Environ. Int.*, vol. 85, no. 4, pp. 206–212, Dec. 2015.
- [28] N. Ribas-Fitó, M. Torrent, D. Carrizo, J. Júlvez, J. O. Grimalt, and J. Sunyer, "Exposure to Hexachlorobenzene during Pregnancy and Children's Social Behavior at 4 Years of Age," *Environ. Health Perspect.*, vol. 115, no. 3, pp. 447–450, Mar. 2007.
- [29] P. Sly *et al.*, "Polychlorinated biphenyls but not chlorinated pesticides are associated with externalizing behaviour in adolescents," *Organohalogen Compd.*, vol. 74, pp. 1285–1287, 2012.
- [30] O. Faroon and P. Ruiz, "Polychlorinated biphenyls: New evidence from the last decade," *Toxicol. Ind. Health*, vol. 32, no. 11, pp. 1825–1847, Nov. 2016.
- [31] A. Chen *et al.*, "Prenatal Polybrominated Diphenyl Ether Exposures and Neurodevelopment in U.S. Children through 5 Years of Age: The HOME Study," *Environ. Health Perspect.*, vol. 122, no. 8, pp. 856–862, Aug. 2014.
- [32] J. B. Herbstman *et al.*, "Prenatal Exposure to PBDEs and Neurodevelopment," *Environ. Health Perspect.*, vol. 118, no. 5, pp. 712–719, May 2010.
- [33] M. H. Harris *et al.*, "Prenatal and childhood exposure to per- and polyfluoroalkyl substances (PFASs) and child cognition," *Environ. Int.*, vol. 115, pp. 358–369, Jun. 2018.
- [34] Y. Wang *et al.*, "Prenatal exposure to perfluoroalkyl substances and children's IQ: The Taiwan maternal and infant cohort study," *Int. J. Hyg. Environ. Health*, vol. 218, no. 7, pp. 639–644, Oct. 2015.
- [35] Z. Liew, H. Goudarzi, and Y. Oulhote, "Developmental Exposures to Perfluoroalkyl Substances (PFASs): An Update of Associated Health Outcomes," *Curr. Environ. Heal. Reports*, vol. 5, no. 1, pp. 1–19, Mar. 2018.
- [36] S. M. Engel *et al.*, "Prenatal Phthalate Exposure Is Associated with Childhood Behavior and Executive Functioning," *Environ. Health Perspect.*, vol. 118, no. 4, pp. 565–571, Apr. 2010.
- [37] R. M. Whyatt *et al.*, "Maternal Prenatal Urinary Phthalate Metabolite Concentrations and Child Mental, Psychomotor, and Behavioral Development at 3 Years of Age," *Environ. Health Perspect.*, vol. 120, no. 2, pp. 290–295, Feb. 2012.
- [38] T. Achenbach and L. Rescorla, *Manual for the ASEBA Preschool Forms & Profiles*. Burlington: University of Vermont, Research Center for Children, Youth & Families, 2001.
- [39] B.-N. Kim *et al.*, "Phthalates Exposure and Attention-Deficit/Hyperactivity Disorder in School-Age Children," *Biol. Psychiatry*, vol. 66, no. 10, pp. 958–963, Nov. 2009.
- [40] A. Miodovnik *et al.*, "Endocrine disruptors and childhood social impairment," *Neurotoxicology*, vol. 32, no. 2, pp. 261–267, Mar. 2011.
- [41] M. M. Téllez-Rojo *et al.*, "Prenatal urinary phthalate metabolites levels and neurodevelopment in children at two and three years of age," *Sci. Total Environ.*, vol. 461–462, pp. 386–390, Sep. 2013.
- [42] R. W. Kobrosly *et al.*, "Prenatal Phthalate Exposures and Neurobehavioral Development Scores in Boys and Girls at 6–10 Years of Age," *Environ. Health Perspect.*, vol. 122, no. 5, pp. 521–528, May 2014.
- [43] C. Philippat *et al.*, "Prenatal exposure to nonpersistent endocrine disruptors and behavior in boys at 3 and 5 years," *Environ. Health Perspect.*, vol. 125, no. 9, pp. 1–9, 2017.
- [44] M. Ejaredar, E. C. Nyanza, K. Ten Eycke, and D. Dewey, "Phthalate exposure and childrens neurodevelopment: A systematic review," *Environ. Res.*, vol. 142, pp. 51–60, Oct. 2015.
- [45] C. Philippat, D. Bennett, A. M. Calafat, and I. H. Picciotto, "Exposure to select phthalates and phenols through use of personal care products among Californian adults and their children," *Environ. Res.*, vol. 140, pp. 369–376, Jul. 2015.
- [46] M. Casas *et al.*, "Exposure to bisphenol A during pregnancy and child neuropsychological development in the INMA-Sabadell cohort," *Environ. Res.*, vol. 142, pp. 671–679, Oct. 2015.
- [47] F. Perera *et al.*, "Prenatal Bisphenol A Exposure and Child Behavior in an Inner-City Cohort," *Environ. Health Perspect.*, vol. 120, no. 8, pp. 1190–4, Aug. 2012.
- [48] S. F. Evans *et al.*, "Prenatal bisphenol A exposure and maternally reported behavior in boys and girls," *Neurotoxicology*, vol.

45, no. 5, pp. 91–99, Dec. 2014.

- [49] K. G. Harley *et al.*, “Prenatal and early childhood bisphenol A concentrations and behavior in school-aged children,” *Environ. Res.*, vol. 126, pp. 43–50, Oct. 2013.
- [50] E. L. Roen *et al.*, “Bisphenol A exposure and behavioral problems among inner city children at 7–9 years of age,” *Environ. Res.*, vol. 142, pp. 739–745, Oct. 2015.
- [51] J. M. Braun *et al.*, “Impact of Early-Life Bisphenol A Exposure on Behavior and Executive Function in Children,” *Pediatrics*, vol. 128, no. 5, pp. 873–882, Nov. 2011.
- [52] J. M. Braun, “Early-life exposure to EDCs: role in childhood obesity and neurodevelopment,” *Nat. Rev. Endocrinol.*, vol. 13, no. 3, pp. 161–173, Mar. 2017.
- [53] D. Nakiwala *et al.*, “In-utero exposure to phenols and phthalates and the intelligence quotient of boys at 5 years,” *Environ. Heal. A Glob. Access Sci. Source*, vol. 17, no. 1, pp. 1–11, 2018.
- [54] T. Etzel *et al.*, “Prenatal urinary triclosan concentrations and child neurobehavior,” *Environ. Int.*, vol. 114, pp. 152–159, May 2018.
- [55] B. Eskenazi *et al.*, “Organophosphate Pesticide Exposure and Neurodevelopment in Young Mexican-American Children,” *Environ. Health Perspect.*, vol. 115, no. 5, pp. 792–798, May 2007.
- [56] Y. Wang *et al.*, “Prenatal and postnatal exposure to organophosphate pesticides and childhood neurodevelopment in Shandong, China,” *Environ. Int.*, vol. 108, pp. 119–126, Nov. 2017.
- [57] P. Liu, C. Wu, X. Chang, X. Qi, M. Zheng, and Z. Zhou, “Adverse Associations of both Prenatal and Postnatal Exposure to Organophosphorous Pesticides with Infant Neurodevelopment in an Agricultural Area of Jiangsu Province, China,” *Environ. Health Perspect.*, vol. 124, no. 10, pp. 1637–1643, Oct. 2016.
- [58] M. A. Furlong *et al.*, “Prenatal exposure to organophosphorus pesticides and childhood neurodevelopmental phenotypes,” *Environ. Res.*, vol. 158, pp. 737–747, Oct. 2017.
- [59] K. Khan *et al.*, “Manganese Exposure from Drinking Water and Children’s Classroom Behavior in Bangladesh,” *Environ. Health Perspect.*, vol. 119, no. 10, pp. 1501–1506, Oct. 2011.
- [60] S. Yousef, A. Adem, T. Zoubeidi, M. Kosanovic, A. A. Mabrouk, and V. Eapen, “Attention Deficit Hyperactivity Disorder and Environmental Toxic Metal Exposure in the United Arab Emirates,” *J. Trop. Pediatr.*, vol. 57, no. 6, pp. 457–460, Dec. 2011.
- [61] C. Salustri *et al.*, “Is cognitive function linked to serum free copper levels? A cohort study in a normal population,” *Clin. Neurophysiol.*, vol. 121, no. 4, pp. 502–507, Apr. 2010.
- [62] F. Harari, M. Bottai, E. Casimiro, B. Palm, and M. Vahter, “Exposure to Lithium and Cesium Through Drinking Water and Thyroid Function During Pregnancy: A Prospective Cohort Study,” *Thyroid*, vol. 25, no. 11, pp. 1199–1208, Nov. 2015.
- [63] K. Polanska *et al.*, “Selenium status during pregnancy and child psychomotor development—Polish Mother and Child Cohort study,” *Pediatr. Res.*, vol. 79, no. 6, pp. 863–869, Jun. 2016.
- [64] J. Al-Dahhan, “Effect of salt supplementation of newborn premature infants on neurodevelopmental outcome at 10–13 years of age,” *Arch. Dis. Child. - Fetal Neonatal Ed.*, vol. 86, no. 2, p. 120F–123, Mar. 2002.
- [65] A. Modabbernia, M. Arora, and A. Reichenberg, “Environmental exposure to metals, neurodevelopment, and psychosis,” *Curr. Opin. Pediatr.*, vol. 28, no. 2, pp. 243–249, Apr. 2016.
- [66] M. B. Hopson, A. Margolis, V. Rauh, and J. Herbstman, “Impact of the home environment on the relationship between prenatal exposure to environmental tobacco smoke and child behavior,” *Int. J. Child Health Hum. Dev.*, vol. 9, no. 4, pp. 453–464, 2016.
- [67] T. T. Luk, M. P. Wang, Y. N. Suen, D. S.-Q. Koh, T. H. Lam, and S. S.-C. Chan, “Early childhood exposure to secondhand smoke and behavioural problems in preschoolers,” *Sci. Rep.*, vol. 8, no. 1, p. 15434, Dec. 2018.
- [68] K. Polanska *et al.*, “Environmental Tobacco Smoke Exposure during Pregnancy and Child Neurodevelopment,” *Int. J. Environ. Res. Public Health*, vol. 14, no. 7, p. 796, Jul. 2017.
- [69] K. Polańska, J. Jurewicz, and W. Hanke, “Smoking and alcohol drinking during pregnancy as the risk factors for poor child neurodevelopment – A review of epidemiological studies,” *Int. J. Occup. Med. Environ. Health*, vol. 28, no. 3, pp. 419–443, May 2015.
- [70] O. Robinson and M. Vrijheid, “The Pregnancy Exposome,” *Curr. Environ. Heal. reports*, vol. 2, no. 2, pp. 204–213, 2015.
- [71] L. S. Haug *et al.*, “In-utero and childhood chemical exposome in six European mother-child cohorts,” *Environ. Int.*, vol. 121, no. June, pp. 751–763, Dec. 2018.

- [72] C. P. Wild, "Complementing the Genome with an 'Exposome': The Outstanding Challenge of Environmental Exposure Measurement in Molecular Epidemiology," *Cancer Epidemiol. Biomarkers Prev.*, vol. 14, no. 8, pp. 1847–1850, Aug. 2005.
- [73] "HELIX Project Website." [Online]. Available: <https://www.projecthelix.eu/>.
- [74] M. Vrijheid *et al.*, "The Human Early-Life Exposome (HELIX): Project Rationale and Design," *Environ. Health Perspect.*, vol. 122, no. 6, pp. 535–544, Jun. 2014.
- [75] L. Maitre *et al.*, "Cohort Profile: the Human Early Life Exposome (HELIX) study - A European Population-Based Exposome Cohort," *BMJ Open*, no. 1, pp. 1–17, 2018.
- [76] R. Goodman, "The Strengths and Difficulties Questionnaire: a research note.," *J. Child Psychol. Psychiatry*, vol. 38, no. 5, pp. 581–6, 1997.
- [77] R. Goodman and S. Scott, "Comparing the Strengths and Difficulties Questionnaire and the Child Behavior Checklist: is small beautiful?," *J. Abnorm. Child Psychol*, vol. 27, no. 1, pp. 17–24, 1999.
- [78] U. Held *et al.*, "Methods for Handling Missing Variables in Risk Prediction Models," *Am. J. Epidemiol.*, vol. 184, no. 7, pp. 545–551, Oct. 2016.
- [79] S. Jolani, T. P. A. A. Debray, H. Koffijberg, S. van Buuren, and K. G. M. M. Moons, "Imputation of systematically missing predictors in an individual participant data meta-analysis: A generalized approach using MICE," *Stat. Med.*, vol. 34, no. 11, pp. 1841–1863, May 2015.
- [80] I. R. White, P. Royston, and A. M. Wood, "Multiple imputation using chained equations: Issues and guidance for practice," *Stat. Med.*, vol. 30, no. 4, pp. 377–399, Feb. 2011.
- [81] S. van Buuren and K. Groothuis-Oudshoorn, "mice: Multivariate Imputation by Chained Equations in R," *J. Stat. Softw.*, vol. 45, no. 3, 2011.
- [82] G. Vink and S. van Buuren, "mice: An approach to sensitivity analysis." [Online]. Available: [http://www.gerkovink.com/miceVignettes/Sensitivity\\_analysis/Sensitivity\\_analysis.html](http://www.gerkovink.com/miceVignettes/Sensitivity_analysis/Sensitivity_analysis.html). [Accessed: 16-Nov-2018].
- [83] G. Vink and S. van Buuren, "mice: Passive imputation and Post-processing." [Online]. Available: [https://www.gerkovink.com/miceVignettes/Passive\\_Post\\_processing/Passive\\_imputation\\_post\\_processing.html](https://www.gerkovink.com/miceVignettes/Passive_Post_processing/Passive_imputation_post_processing.html). [Accessed: 16-Nov-2018].
- [84] K. M. O'Brien, K. Upson, and J. P. Buckley, "Lipid and Creatinine Adjustment to Evaluate Health Effects of Environmental Exposures," *Curr. Environ. Heal. Reports*, vol. 4, no. 1, pp. 44–50, Mar. 2017.
- [85] M. Mortamais *et al.*, "Correcting for the influence of sampling conditions on biomarkers of exposure to phenols and phthalates: a 2-step standardization method based on regression residuals," *Environ. Heal.*, vol. 11, no. 1, p. 29, Dec. 2012.
- [86] K. J. Rothman, S. Greenland, and T. L. Lash, *Modern epidemiology*. LWW, 2008.
- [87] J. M. Hilbe, *Negative Binomial Regression*, Second Edi. Cambridge University Press, 2011.
- [88] M.-X. Li, J. M. Y. Yeung, S. S. Cherny, and P. C. Sham, "Evaluating the effective numbers of independent tests and significant p-value thresholds in commercial genotyping arrays and public imputation reference datasets," *Hum. Genet.*, vol. 131, no. 5, pp. 747–756, May 2012.
- [89] L. Thabane *et al.*, "A tutorial on sensitivity analyses in clinical trials: the what, why, when and how," *BMC Med. Res. Methodol.*, vol. 13, no. 1, p. 92, Dec. 2013.
- [90] K. J. Rothman, "No adjustments are needed for multiple comparisons.," *Epidemiology*, vol. 1, no. 1, pp. 43–6, Jan. 1990.
- [91] Y. Kim *et al.*, "Prenatal Exposure to Phthalates and Infant Development at 6 Months: Prospective Mothers and Children's Environmental Health (MOCEH) Study," *Environ. Health Perspect.*, vol. 119, no. 10, pp. 1495–1500, Oct. 2011.
- [92] Y. Lien *et al.*, "Prenatal exposure to phthalate esters and behavioral syndromes in children at 8 years of age: Taiwan Maternal and Infant Cohort Study," *Env. Heal. Perspect*, vol. 123, no. 1, pp. 95– 100, 2015.
- [93] L. M. Gaetke, H. S. Chow-Johnson, and C. K. Chow, "Copper: toxicological relevance and mechanisms," *Arch. Toxicol.*, vol. 88, no. 11, pp. 1929–1938, Nov. 2014.
- [94] Centres for Disease Control and Prevention, "Fourth National Report on Human Exposure to Environmental Chemicals. Updated Tables. Volume One," 2018.
- [95] J. Vukelic *et al.*, "Variations of serum copper values in pregnancy," *Srp. Arh. Celok. Lek.*, vol. 140, no. 1–2, pp. 42–46, 2012.
- [96] F. Elbaz, S. Zahra, and H. Hanafy, "Magnesium, zinc and copper estimation in children with attention deficit hyperactivity disorder (ADHD)," *Egypt. J. Med. Hum. Genet.*, vol. 18, no. 2, pp. 153–163, Apr. 2017.



- [97] A. Viktorinova, M. Ursinyova, J. Trebaticka, I. Uhnakova, Z. Durackova, and V. Masanova, "Changed Plasma Levels of Zinc and Copper to Zinc Ratio and Their Possible Associations with Parent- and Teacher-Rated Symptoms in Children with Attention-Deficit Hyperactivity Disorder," *Biol. Trace Elem. Res.*, vol. 169, no. 1, pp. 1–7, Jan. 2016.
- [98] M. Kicinski *et al.*, "Neurobehavioral function and low-level metal exposure in adolescents," *Int. J. Hyg. Environ. Health*, vol. 218, no. 1, pp. 139–146, Jan. 2015.
- [99] G. M.-M. Bumoko *et al.*, "Lower serum levels of selenium, copper, and zinc are related to neuromotor impairments in children with konzo," *J. Neurol. Sci.*, vol. 349, no. 1–2, pp. 149–153, Feb. 2015.
- [100] K. J. Pieper *et al.*, "Evaluating Water Lead Levels During the Flint Water Crisis," *Environ. Sci. Technol.*, vol. 52, no. 15, pp. 8124–8132, Aug. 2018.
- [101] Centers for Disease Control and Prevention, "Biomonitoring Summary for Non-Dioxin-Like Polychlorinated Biphenyls," *National Biomonitoring Program*, 2016. [Online]. Available: [https://www.cdc.gov/biomonitoring/NDL-PCBs\\_BiomonitoringSummary.html](https://www.cdc.gov/biomonitoring/NDL-PCBs_BiomonitoringSummary.html). [Accessed: 15-Dec-2018].
- [102] H. Zhang *et al.*, "Prenatal PBDE and PCB Exposures and Reading, Cognition, and Externalizing Behavior in Children," *Environ. Health Perspect.*, vol. 125, no. 4, pp. 746–752, Apr. 2017.
- [103] S. K. Sagiv *et al.*, "Sociodemographic and Perinatal Predictors of Early Pregnancy Per- and Polyfluoroalkyl Substance (PFAS) Concentrations," *Environ. Sci. Technol.*, vol. 49, no. 19, pp. 11849–11858, Oct. 2015.
- [104] J. M. Braun *et al.*, "Gestational exposure to endocrine-disrupting chemicals and reciprocal social, repetitive, and stereotypic behaviors in 4- and 5-year-old children: The HOME study," *Environ. Health Perspect.*, vol. 122, no. 5, pp. 513–520, 2014.
- [105] J. Barrera-Gómez *et al.*, "A systematic comparison of statistical methods to detect interactions in exposome-health associations," *Environ. Heal. A Glob. Access Sci. Source*, vol. 16, no. 1, pp. 1–13, 2017.

=====

Pages I – XII were removed

=====